

## Different effects of a bradykinin B<sub>1</sub> selective antagonist and B<sub>2</sub> selective antagonist on Mas-related G protein-coupled receptors in rat mast cells, and inhibitory effects of lithium carbonate

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**Abstract** Mas-related G protein-coupled receptors (MRGPRs) in mast cells are the targets of pseudoallergic drug reactions. Bradykinin (BK) and its analogues exert agonistic effects on mast cells; however, it currently remains unclear whether these effects depend on the activation of MRGPRs. In the present study, we examined the effects of the BK analogues on rat mast cells in order to clarify the influence of amino acid sequences on mast cell activation via MRGPRs. The B<sub>1</sub> selective agonist released histamine, whereas the B<sub>1</sub> selective antagonist did not inhibit histamine release induced by BK and the B<sub>1</sub> agonist. The B<sub>2</sub> antagonist did not exert inhibitory effects. The B<sub>2</sub> antagonist exerted agonistic effects, whereas the B<sub>1</sub> selective antagonist did not. The agonistic effects of the B<sub>1</sub> agonist and B<sub>2</sub> antagonist depended on the number of cationic amino acids in the amino acid sequence. The effects of lithium carbonate were investigated, and it was found to inhibit histamine release induced by the B<sub>2</sub> antagonist, suggesting that the B<sub>2</sub> antagonist is a MRGPR activator. The MRGPRs activated by selective B<sub>1</sub> agonist and selective B<sub>2</sub> antagonist are one of the pathways that induce pseudoallergic drug reactions. Lithium carbonate will contribute to the identification of candidates that control pseudoallergic drug reactions.

**Key words:** Mas-related G protein-coupled receptor, mast cell activation, bradykinin, bradykinin agonist, bradykinin antagonist

### Introduction

Mast cells express Mas-related G protein-coupled receptors (MRGPRs), mouse *Mrgprb2*, rat *MrgprC*, and human MRGPRX2, which are the targets of allergic, anaphylactic, and pseudoallergic drug reactions<sup>1-3</sup>. Previous studies demonstrated that substance P (SP)<sup>4</sup> and bradykinin (BK)<sup>5</sup> released histamine from rat mast cells in an IgE-independent

manner prior to the discovery of MRGPRs in mast cells. BK has 2 classes of receptors, BK B<sub>1</sub> receptor (B<sub>1</sub>) and BK B<sub>2</sub> receptor (B<sub>2</sub>). Devillier et al. reported that B<sub>2</sub> selective antagonists were potent releasers of histamine from rat mast cells and did not exert any antagonistic effects<sup>6</sup>. Azimi et al. recently showed that the neurokinin-1 antagonist aprepitant did not inhibit the activation of human MRGPRX2, but tripeptide

QWF (Boc-Gln-D-Trp (Formyl)-Phe benzyl ester trifluoroacetate) inhibited  $\beta$ -hexosaminidase release from a human mast cell line activated by SP after blocking the binding of SP to MRGPRX2<sup>7</sup>. On the other hand, the B2 antagonist icatibant did not exert inhibitory effects on human mast cell line, suggesting that it caused pseudoallergic drug reaction on MRGPRX2<sup>8,9</sup>. In the present study, we examined the effects of a B1 selective agonist, B1 selective antagonist, and B2 selective antagonist (Table 1) on rat peritoneal mast cells in order to clarify the influence of amino acid sequences of bradykinin analogues on mast cell activation via MRGPRs. The effects of lithium carbonate, which is an inhibitor of histamine release induced by compound 48/80, SP, BK, mastoparan and melittin<sup>10</sup> were also investigated.

## Materials and Methods

### Chemicals

BK acetate salt, the B1 selective agonist [Lys-des-Arg<sup>9</sup>]-BK, and lithium carbonate (Li<sub>2</sub>CO<sub>3</sub>) were purchased from Sigma-Aldrich (U.S.A.). The B1 selective antagonist des-Arg<sup>9</sup>-[Leu<sup>8</sup>]-BK and B2 selective antagonist [Thi<sup>5,8</sup>-D-Phe<sup>7</sup>]-BK were from Peptide Institute Inc. (Osaka, Japan). Thi indicates thienylalanine.

**Table 1** Amino acid sequences of bradykinin, the B1 selective agonist, B1 selective antagonist, and B2 selective antagonist used in the present study.

Bradykinin and analogues	Amino acid sequence
Bradykinin	Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe-Arg
B1 selective agonist	Lys-Arg-Pro-Pro-Gly-Phe-Ser-Pro-Phe
B1 selective antagonist	Arg-Pro-Pro-Gly-Phe-Ser-Pro-Leu
B2 selective antagonist	Arg-Pro-Pro-Gly-Thi-Ser-D-Phe-Thi-Arg

\*Thi indicates thienylalanine.

### Assay of histamine release

Histamine release from rat peritoneal mast cells was

measured as described previously<sup>11</sup>. Mast cells from the peritoneal cavity of male Sprague Dawley rats (300-450 g) were purified using Percoll and suspended in HEPES-buffered Tyrode's solution (137 mM NaCl, 2.7 mM KCl, 12 mM HEPES, 1 mM MgCl<sub>2</sub>, 0.3 mM CaCl<sub>2</sub>, 5.6 mM dextrose, and 0.03% bovine serum albumin, pH 7.4). Mast cells were preincubated with the B1 selective antagonist or Li<sub>2</sub>CO<sub>3</sub> in HEPES-buffered Tyrode's solution at 37°C for 5 min, and 50  $\mu$ M BK or 50  $\mu$ M of the B2 selective antagonist was then added. After a 5 min incubation, the amount of histamine released was analyzed using a fluorometric assay<sup>12</sup>.

Mast cells were also preincubated with 3 mM Li<sub>2</sub>CO<sub>3</sub> at 37°C for 5 min before the incubation with 50  $\mu$ M of the B2 selective antagonist for 5 min.

### Animal studies

All animal studies were previously approved by the Animal Care Committee of Shujitsu University (#018-001 and #18-002) and conducted in accordance with the Principles of Laboratory Animal Care (NIH Publication #85-23). Sprague-Dawley male rats were used in animal studies.

### Statistical analysis

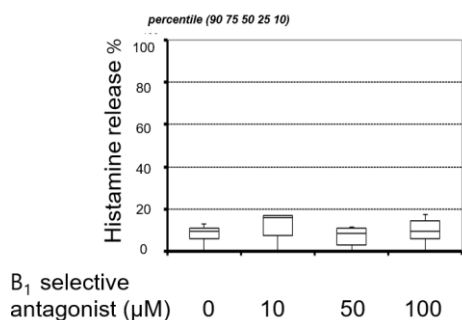
Values are box-and-whisker plots of the replicated experiments. Statistical analyses were performed using repeated measures ANOVA with Dunnett's test employing P=0.05 as the upper limit of significance.

## Results

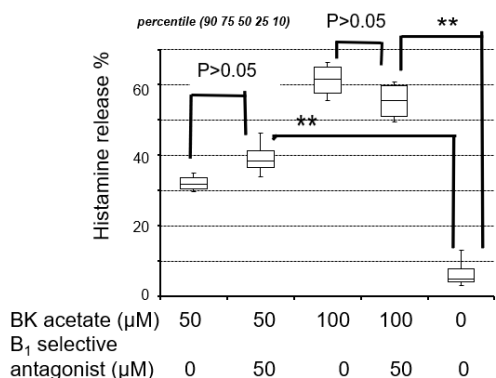
### 1. The B1 selective antagonist did not inhibit histamine release induced by BK.

At concentrations at 10, 50, and 100  $\mu$ M, the B1 selective antagonist did not enhance spontaneous histamine release (Fig. 1). The effects of the B1 selective antagonist on histamine release induced by 50 and 100  $\mu$ M BK were examined. The B1 selective

antagonist did not inhibit histamine release induced by 50 or 100  $\mu\text{M}$  BK (Fig. 2). Therefore, we examined the effects of the B1 selective antagonist on histamine release induced by the B1 selective agonist. The B1 selective antagonist did not enhance or inhibit histamine release induced by the B1 selective agonist (Fig. 3).



**Fig. 1 Histamine release induced by the B1 selective antagonist.** Mast cells were preincubated with 10, 50, and 100  $\mu\text{M}$  of the B1 selective antagonist in HEPES-buffered Tyrode's solution at 37°C for 10 min. The amount of histamine released was analyzed, n=6, Dunnett's analysis, \*\*P<0.01 Repeated measures ANOVA.

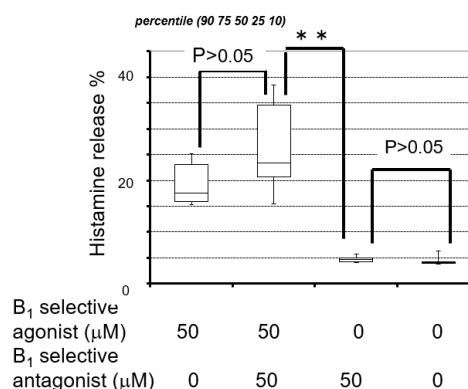


**Fig. 2 Effects of the B1 selective antagonist on histamine release induced by 50  $\mu\text{M}$  BK.** Mast cells were preincubated with 50  $\mu\text{M}$  of the B1 selective antagonist in HEPES-buffered Tyrode's solution at 37°C for 5 min, and 50  $\mu\text{M}$  BK was then added. After a 5 min incubation, the amount of histamine released was analyzed, n=4, Dunnett's analysis, \*\*P<0.01 Repeated measures ANOVA.

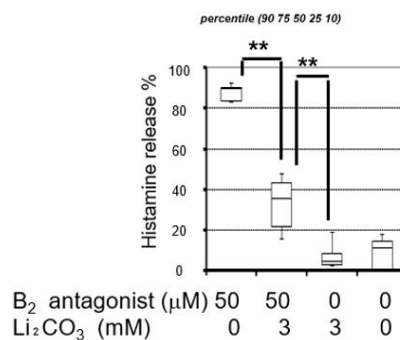
## 2. The B2 selective antagonist released histamine from rat mast cells, and lithium carbonate inhibited this release.

Since the B2 selective antagonist induced the release of histamine, it was not possible to examine the inhibitory effects of the B2 selective antagonist on histamine release induced by BK (data not shown). Lithium carbonate dose-dependently inhibited

histamine release induced by BK, with  $\text{IC}_{50}$  of 1.50  $\text{mM}^{(10)}$ , therefore, the effects of lithium carbonate on histamine release induced by the B2 selective antagonist were examined. Lithium carbonate inhibited histamine release induced by the B2 selective antagonist, with an  $\text{IC}_{50}$  of approximately 2.25  $\text{mM}$  (Fig. 4).



**Fig. 3 Effects of the B1 selective antagonist on histamine release induced by 50  $\mu\text{M}$  of the B1 selective agonist.** Mast cells were preincubated with 50  $\mu\text{M}$  of the B1 selective antagonist in HEPES-buffered Tyrode's solution at 37°C for 5 min, and 50  $\mu\text{M}$  of the B1 selective agonist was then added. After a 5 min incubation, the amount of histamine released was analyzed, n=6, Dunnett's analysis, \*\*P<0.01 Repeated measures ANOVA



**Fig. 4 Effects of 3 mM lithium carbonate on histamine release induced by 50  $\mu\text{M}$  of the B2 selective antagonist.** Mast cells were preincubated with 3.0 mM lithium carbonate in HEPES-buffered Tyrode's solution at 37°C for 5 min, and 50  $\mu\text{M}$  of the B2 antagonist was then added. After a 5 min incubation, the amount of histamine released was analyzed, n=6, Dunnett's analysis, \*\*P<0.01 Repeated measures ANOVA.

## Discussion

Before the presence of MRGPRs on mast cells, which play roles in mast cell activation, was demonstrated, histamine release by kinins was either a non-specific effect not mediated by receptors or, if receptors were

involved, these sites were suggested to differ from other kinin receptors (i.e. B<sub>1</sub> or B<sub>2</sub>)<sup>6)</sup>. SP and BK are kinins that induce the release of histamine, and two B<sub>1</sub> antagonists, des-Arg<sup>10</sup>-[Leu<sup>9</sup>]kallidin and des-Arg<sup>9</sup>-[Leu<sup>8</sup>]BK, were shown to promote histamine release and acted only as agonists<sup>6)</sup>. The order of potency of these analogues was as follows: [Thi<sup>6,9</sup>,D-Phe<sup>8</sup>]kallidin > [Thi<sup>5,8</sup>,D-Phe<sup>7</sup>]BK > des-Arg<sup>10</sup>-[Leu<sup>9</sup>]kallidin > des-Arg<sup>9</sup>-[Leu<sup>8</sup>]BK<sup>6)</sup>. In the present study, we examined the effects of a B<sub>1</sub> selective agonist, B<sub>1</sub> selective antagonist, and B<sub>2</sub> selective antagonist. These peptides did not exert inhibitory effects. The agonistic effects observed depended on the number of cationic amino acids in the sequence, suggesting that at least 2 positively charged amino acids are needed to induce MRGPR activation (Table 1). The B<sub>1</sub> selective antagonist des-Arg<sup>9</sup>-[Leu<sup>8</sup>]BK did not release histamine, suggesting that one Arg residue is insufficient to activate MRGPRs. Moreover, the B<sub>1</sub> selective antagonist did not inhibit histamine release induced by BK and the B<sub>1</sub> selective agonist. Devillier et al. previously reported that the agonistic effects of the B<sub>1</sub> antagonist des-Arg<sup>9</sup>-[Leu<sup>8</sup>]BK were weak<sup>6)</sup>, which is inconsistent with the present results. Under our assay conditions that the B<sub>1</sub> selective antagonist des-Arg<sup>9</sup>-[Leu<sup>8</sup>]BK did not affect spontaneous histamine release, we were able to examine the effects of the B<sub>1</sub> selective antagonist on histamine release induced by BK and the B<sub>1</sub> agonist (Figs. 1, 2, and 3). In the present study, the B<sub>2</sub> antagonist [Thi<sup>5,8</sup>,D-Phe<sup>7</sup>]BK stimulated histamine release. Therefore, the inhibitory effects of lithium carbonate were investigated (Fig. 4). Lithium carbonate is one of the treatments for bipolar disorder; however, its mechanisms of action remain unclear. We previously reported that lithium carbonate inhibited histamine release from rat mast cells induced by compound 48/80, SP, BK, mastoparan, melittin, and GlcNAc-GlcNAc specific lectin *Datura stramonium* agglutinin,

suggesting that it is one of the inhibitors of MRGPRs<sup>10)</sup>. Since lithium carbonate inhibited histamine release induced by the B<sub>2</sub> antagonist, the B<sub>2</sub> antagonist appears to stimulate MRGPRs.

The high activity levels of BK and its analogues suggested that their potencies to induce the release of histamine from rat mast cells increases in parallel with the number of positively charged amino acids (Lys and Arg), and the N-terminal arginine appears to be important<sup>13)</sup>. On the other hand, peptide sequences are not necessary to activate the MRGPRs of mast cells by BK because icatibant, a B<sub>2</sub> antagonist with no peptide chain, promoted mast cell degranulation via MRGPRX2 and induced a pseudoallergic drug reaction<sup>8,9)</sup>. It is important to note that the degranulation and intracellular calcium ion increases induced by icatibant were not observed in MRGPRX2 knockdown culture cells.

Mast cells are activated in not only IgE-dependent, but also IgE-independent manners. The IgE-independent pathway is due to anaphylactic and pseudoallergic drug reactions because many polycationic chemicals stimulate this pathway. Regarding many polycationic chemicals, the IgE-independent pathway appears to have been misunderstood as a non-specific effect not mediated by receptors for a long time. MRGPRs are predominantly expressed in sensory neurons and mast cells, and are the target of polycationic chemicals to induce pseudoallergic drug reactions<sup>14)</sup>. The selective B<sub>1</sub> agonist and selective B<sub>2</sub> antagonist are also activators. Furthermore, the inhibitory concentration of lithium carbonate on MRGPRs was shown to be close to its clinical and addictive doses<sup>15)</sup>. Lithium carbonate as an inhibitor of mast cells has potential as a candidate to control anaphylactic and pseudoallergic drug reactions.

**Conflicts of Interest:** The authors declare no conflicts

of interest.

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